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(54) Title: STRESS TOLERANT YEAST MUTANTS

(57) Abstract

The invention provides methods and compositions relating to stress tolerant yeasts; in particular, yeast mutants deficient in the expression of functional ATH1 gene product (Ath1p). Such yeast have enhanced tolerance to dehydration and freezing, are able to grow to a higher cell density over a range of fermentable carbon source concentrations, are able to produce and/or tolerate higher levels of ethanol and trehalose. Nucleic acids comprising ATH1 gene sequences are used in hybridization probes and PCR primers, in expression vectors, etc. The invention provides methods for producing a yeast mutant with improved survival ability under stress conditions which involve identifying mutations disrupting ATH1 expression using Ath1-specific reagents or ATH1 hybridization probes or primers.

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STRESS TOLERANT YEAST MUTANTS

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5 any patent issuing on this application.

INTRODUCTIONTechnical Field

The technical field of this invention concerns a specific genetic mutation in
10 yeast which provides enhanced stress tolerance.

Background

The non-reducing disaccharide *O*- α -D-glucopyranosyl-1 \rightarrow 1- α -D-glucopyranoside, commonly known as trehalose, was discovered in 1832 (Wiggers, 1832)
15 in a fungus, *Secale cornutum*. Since then, trehalose has been found in a wide variety of organisms including additional fungi, bacteria, plants, insects and other invertebrates. In *Saccharomyces cerevisiae*, trehalose is one of the major storage carbohydrates, accounting for up to 23% or more of the dry weight of the cells, depending on growth conditions and the stage of life cycle (Elbein, 1974).

20 Trehalose is believed to function in yeast as an energy source in spore germination and as a protecting agent for maintaining structural integrity under environmental stresses such as heat and desiccation (Thevelein, 1984). More recent results, however, indicate that the bulk of trehalose accumulated in yeast under mild heat treatment is not sufficient to account for the acquisition of
25 thermotolerance (Arguelles, 1994; Nwaka et al., 1994; Winkler et al., 1991). The concentration of trehalose in the yeast cell is the result of the activities of the synthesizing bifunctional enzyme trehalose-6-phosphate-synthase/trehalose-6-phosphate phosphatase (Vuorio et al., 1993) and the trehalose hydrolyzing enzymes, e.g. cytosolic neutral trehalase (App and Holzer, 1989) and vacuolar acid
30 trehalase (Mittenbühler and Holzer, 1988). The recently cloned neutral trehalase (NTH) is considered to be the key enzyme responsible for trehalose degradation

in intact yeast cells (Kopp et al., 1993; Wiemken, 1990), however, very little is known about the biological function and possible control mechanisms for vacuolar acid trehalase (ATH). ATH has been shown to be glycosylated (Londesborough and Varimo, 1984; Mittenbühler and Holzer, 1988) and activation is dependent on the *PEP4* gene product, proteinase A (Harris and Cotter, 1987). The physiological role of ATH and the coordination of its function with that of NTH is unknown.

Due to its role in stress protection, trehalose has important commercial applications for the baking and brewing industries (Mansure et al., 1994; Oda et al., 1986; Hino et al., 1990; Gelinas et al., 1989). The synthesis and degradation of trehalose is important in yeast cell physiology at various stages of growth; mobilization of trehalose and the timing of its metabolism are critical for yeast growth and survival.

SUMMARY OF THE INVENTION

The invention provides methods and compositions relating to stress tolerant yeast; in particular, yeast mutants deficient in the expression of functional ATH1 gene product (Ath1p). An exemplary haploid *S. cerevisiae* strain, identified as MDY3, is deposited at the Section of Microbiology, University of California, Davis.

Such yeast have broad industrial application. For example, in the baking industries, the enhanced tolerance to dehydration and freezing make the mutant yeast particularly suited for use in frozen dough and dehydrated yeast products. In brewing, the mutant yeast strains are able to grow to a higher cell density over a range of fermentable carbon source (e.g. glucose) concentrations and are able to produce and/or tolerate higher levels of ethanol. Accordingly, these strains are used to generate higher ethanol concentrations, take fermentation to a greater degree of completion (to make drier wine) and complete fermentation faster. The subject yeast also find use as an improved source of trehalose (trehalose is used commercially as a protectant in food and pharmaceutical processes) and as a source of ethanol as fuel or additive for spirits: e.g. using inexpensive fermentation substrates such as molasses or corn syrup.

The invention also encompasses isolated nucleic acids comprising ATH1 (SEQUENCE ID NO:1) or fragment thereof capable of hybridizing under stringent conditions with ATH1; and in particular, genetic constructs comprising in 5' - 3' orientation, a first ATH1 fragment capable of hybridizing under stringent
5 conditions with ATH1, an intervening sequence, and a second different ATH1 fragment capable of hybridizing under stringent conditions with ATH1.

The invention provides methods for producing a yeast mutant with improved survival ability under stress conditions which involve identifying mutations disrupting ATH1 expression using Ath1-specific reagents or ATH1 hybridization
10 probes or primers.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 shows growth versus percent survival over time following dehydration curves.

15 Figure 2 shows growth versus percent ethanol over glucose concentration curves.

Figure 3 shows growth versus trehalose concentration over glucose concentration curves.

20 Figure 4A shows growth versus percent survival over time following dehydration curves.

Figure 4B shows percent survival versus trehalose concentration over time following dehydration curves for ATH1 mutant yeast.

Figure 4C shows percent survival versus trehalose concentration over time following dehydration curves for wild-type yeast.

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DESCRIPTION OF SPECIFIC EMBODIMENTS

The subject yeast mutants are specifically deficient in the expression of a functional ATH1 gene product. Such a mutant expresses less than half, preferably less than 25%, more preferably less than 10% and more preferably less than 1%
30 of the functional Ath1p expressed by the corresponding wild-type yeast. A variety of genetic mutations yield mutants deficient in the expression of functional ATH1

gene product; preferred mutants have at least one ATH1 allele rendered non-functional (i.e. incapable of generating a functional ATH1 gene product). In one embodiment, the invention provides such mutants wherein said mutant or an ancestor of said mutant was generated by genetically engineering a yeast cell to
5 create a nonfunctional mutation in an Ath1p allele of said yeast cell. A particular exemplary mutant, known herein as MDY3, is on deposit at the University of California, Davis, Section of Microbiology.

Phenotypically, such yeast mutants share a number of characteristics including enhanced survival following dehydration, enhanced ethanol tolerance, and
10 enhanced trehalose production. The post-dehydration survivability enhancement is most apparent as the yeast transit from exponential to stationary growth phases. For cells experiencing dehydration at time points from about 0-10 hours following that transition, and usually from about 1-5 hours following transition, until about 20, often 30, and even 40 or more hours later, the mutant yeast demonstrate a
15 significant enhancement in survivability as compared with the corresponding wild-type yeast. See for example, Figure 1 and Figure 4 A. This enhanced survivability often correlates with enhanced trehalose concentrations; see, Figures 4B and 4C. Mutant survival is generally at least 10%, preferably at least 20%, more preferably at least 50%, more preferably at least 100% (i.e. double) more
20 than the corresponding wild-type yeast during at least one time point.

The subject mutants demonstrate enhanced ethanol tolerance. The mutant yeast are thus able to generate higher ethanol media concentrations than their wild-type counterparts: generally at least 5%, preferably at least 10%, more preferably at least 20% higher ethanol concentration at least one nutrient condition and time
25 point. See for example, Figure 2. The subject mutants also demonstrate enhanced trehalose concentrations as compared with their wild-type counterparts: generally at least 5%, preferably at least 10%, more preferably at least 20% higher trehalose concentration at least one nutrient condition and time point. See for example, Figure 3.

30 The invention provides isolated nucleic acids comprising ATH1 (SEQUENCE ID NO:1) or fragments thereof capable of hybridizing under

stringent conditions with ATH1. The subject nucleic acids are either isolated, partially purified, or recombinant. An "isolated" nucleic acid is present as other than a naturally occurring chromosome or transcript in its natural state and isolated from (not joined in sequence to) at least one nucleotide with which it is normally associated on a natural chromosome; a partially pure nucleic acid constitutes at least about 5%, preferably at least about 30%, and more preferably at least about 90% by weight of total nucleic acid present in a given fraction; and a recombinant nucleic acid is flanked - joined in sequence on at least one side - by at least one nucleotide with which it is not normally associated on a natural chromosome.

10 The subject nucleic acids include ATH1 probes and primers comprising one or more ATH1 fragments capable of hybridizing with ATH1 under stringent conditions, e.g. under stringency conditions characterized by a hybridization buffer comprising 0% formamide in 0.9 M saline/0.09 M sodium citrate (SSC) buffer at a temperature of 37°C and remaining bound when subject to washing at 42°C with the SSC buffer at 37°C. Preferred nucleic acids will hybridize in a hybridization buffer comprising 20% formamide in 0.9 M saline/0.09 M sodium citrate (SSC) buffer at a temperature of 42°C and remaining bound when subject to washing at 42°C with 2 X SSC buffer at 42°C.

20 The subject nucleic acids may be introduced into a variety of genetic constructs, plasmids, vectors and cells. For example, a construct useful in generating ATH1 deletion mutants comprises in 5' - 3' orientation, a first ATH1 fragment thereof capable of hybridizing under stringent conditions with ATH1, an intervening sequence, and a second different ATH1 fragment thereof capable of hybridizing under stringent conditions with ATH1.

25 The invention also provides ATH1 gene products and ATH1 gene product-specific binding agents. ATH1 gene products include ATH1 translation products such as Ath1p (SEQUENCE ID NO:2). Binding agents specific for such gene products are produced or identified by a variety of ways. For example, Ath1p peptides are used as immunogens to generate specific polyclonal or monoclonal antibodies. See, Harlow and Lane (1988) Antibodies, A Laboratory Manual, Cold Spring Harbor Laboratory, for general methods. Other prospective Ath1p-peptide

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specific agents are screened from large libraries of synthetic or natural compounds using any convenient binding assay. Such binding agents are capable of binding an ATH1 gene product with an equilibrium constant at least about 10^7 M^{-1} , preferably at least about 10^8 M^{-1} , more preferably at least about 10^9 M^{-1} .

- 5 The invention provide methods for producing ATH1 yeast mutants with improved survival ability under stress conditions. In general, the methods involve subjecting a population of yeast to stress conditions, detecting in said population a yeast mutant deficient in the expression of functional Ath1p gene product and growing said yeast mutant to obtain yeast with improved survival ability under
- 10 stress conditions. The methods may include subjecting the yeast population to conditions promoting mutation, which may be random (e.g. chemical, uv radiation, etc.) or site-directed mutagenesis conditions, of one or more ATH1 alleles and selection for the mutant genotype. The stress conditions provide a selective growth and/or survival advantage for ATH1 mutants deficient in the expression of
- 15 functional Ath1p gene product (e.g. elevated ethanol media concentration, dehydration, etc.). Targeted mutations are conveniently detected using ATH1 specific oligonucleotide primers or probes, by using Ath1p gene product-specific binding agents (i.e. detecting a deficiency in the expression of functional ATH1 gene product) such as Ath1p specific antibodies, or any other convenient method.
- 20 The following examples are offered by way of illustration and not by way of limitation.

EXAMPLES

Protocol for Figure 1

- 25 Examination of effect of Δath1 mutation on survival following dehydration.
1. Grow SEY6210 or SEY6210 $\Delta\text{ath1}::\text{URA3}$ in YNBD medium or YNBD medium -URA. Subculture into YPD (2% glucose) and monitor growth.
 2. Spin 4 x 5 O.D.s at 14 hour time point and wash in 50 mM MES pH 5.5.
 - 5.5. Resuspend in 0.45 ml 50 mM MES pH 5.5.
 - 30 3. Transfer 0.1 ml to two new microfuge tubes and hold at 24°C .
 4. Read O.D.600 of remaining sample (25 ul in 2 ml).

5. Allow one set of samples to dehydrate in speed vac to constant weight (9 hours). Keep at room temperature 8 days. Rehydrate for 10 minutes in 0.5 ml 50 mM MES prewarmed to 40°C.
6. Pellet one set and freeze at -20°C for determination of carbohydrate.
- 5 7. Dilute remaining two samples by adding 400 ul of MES. Freeze one sample by cooling to 4°C @ 4°C/min, 2°C @ 1°C/min, -20°C @ 0.5°C/min and holding at -20°C for 10 min. Thaw rapidly in water bath at 30°C.
8. Hold remaining sample at room temperature for control and plate during freezing of experimental cells.
- 10 9. Dilute cells in 50 mM MES. Plate (50 ul) in triplicate and compare frozen and dehydrated cells to control cells for viability count:
Dehydrated: 1:1000K--3; 1:100K--3; 1:10K--3; 1:100--3; 1:10--3; 1:1--3
Frozen, control: 1:1000K--3; 1:100K--3; 1:10K--3
- 15 Protocol for Figures 2 and 3
Examine growth of Δ ath1 strain in varying glucose concentrations.
 1. Grow SEY6210 and dath1 in YPD (2%). Subculture and grow to O.D. = 3.0.
 2. Inoculate YPD (5 ml) having various concentrations of glucose (0-40%).
- 20 Start cultures at O.D. = 0.125.
 3. Check O.D. after 24, 48 and 72 hours (50 ul in 2.45 ml).
 4. Remove samples for ethanol, glucose and trehalose analysis. Remove 25 O.D.s of cells from each glucose concentration at the 48 hour time point and spin. Remove supernatant (1.0 ml) and freeze for analysis of glucose and ethanol
- 25 levels. Wash pellet in 1X YNB and freeze for analysis of trehalose.
- Protocol for figure 4
Examine effect of Δ ath1 mutation on survival following drying (growth in minimal medium).
 - 30 1. Grow SEY6210 or SEY6210 dath1::URA3 in 100 ml YNBD medium (plus URA for both) to respiratory or stationary phase.

2. Spin 4 x 5 O.D.s [for trehalose assay: spin an additional 10 O.D.s] and wash in 1X YNB. Resuspend in 1.5 ml 1X YNB [for trehalose assay: wash twice, R/S in 100 ul and read O.D. of 5 ul. Freeze remainder].
3. Transfer 0.47 ml to three new microfuge tubes.
- 5 4. Read O.D.600 of remaining sample (25 ul in 1 ml).
5. Pellet two sets, remove supernatant and R/S in 120 ul of 1X YNB. Read O.D.600 of 5 ul. Transfer 100 ul to new tube, weigh and place in speed-vac. Lyophilize until constant weight (3-5 hours). Keep at room temperature for 33 days. Rehydrate one set in 1X YNB prewarmed to 40°C (0.47
- 10 ml) and plate in dilutions. Other set is a back-up.
6. Hold one set at room temperature for controls and plate during dehydration of experimental cells.
7. Dilute in 1X YNB. Plate (50 ul) in triplicate and compare dehydrated cells to control cells for viability count.

15

Strains and media: the *Escherichia coli* strains used in this study were MC1061 F⁻ *hsdR hsdM⁺ araD139 Δ(araABOIC-leu)7679 ΔlacX74 galU galK rpsL* (Casadaban and Cohen, 1980) and DH5a F⁻ *ø80dlacZΔM15 Δ(lacZYA-argF)U169 deoR recA1 endA1 hsdR17 supE44 l⁻ thi-1 gyrA96 relA1*. The yeast strains used

20 were SEY6210 *MATa ura3-52 leu2-3,112 his3-Δ200 trp1-Δ901 lys2-801 suc2Δ9* and SEY6211 *MATa ura3-52 leu2-3,112 his3-Δ200 trp1-Δ901 ade2-101 suc2Δ9*. Standard methods were used to construct yeast strain MDY3 *ura3-52 leu2-3,112 his3-Δ200 trp1-Δ901 lys2-801 suc2Δ9 Δath1::URA3*. Standard yeast (Sherman et al., 1979; Wickerham, 1946) and *E. coli* media (Miller, 1972) were used and

25 supplemented as needed.

Reagents. YNB, Bacto Tryptone, Bacto Peptone, Bacto Yeast Extract and Bacto Agar were from Difco Laboratories (Detroit, Mich.). DNA restriction and modifying enzymes were from New England Biolabs, Inc. (Beverly, Mass.), and Boehringer Mannheim Biochemicals (Mannheim, Germany). Hybond N⁺

30 membranes for Southern and Northern (RNA) blots, [α -³²P]dCTP (3,000 Ci/mmol), and [³⁵S]dATPaS (>1,000 Ci/mmol) were from Amersham Buchler

(Braunschweig, Germany). Random priming materials and additional enzymes were from United States Biochemical Corp. (Cleveland, Ohio). Biochemical reagents were from Sigma (Deisenhofen, Germany).

Enzymatic overlay assay. Preparation and transformation of competent
5 yeast cells by the lithium-acetate method was carried out as described by Ito (Ito et al., 1983). Yeast colonies transformed with DNA from a genomic plasmid library were replica-plated on YP (1% Bacto Yeast Extract, 2% Bacto Peptone) plates with 2% fructose as the carbohydrate source. After growth for 2 days at 30°C, 10 ml of an overlay-assay-mix that measures secreted ATH activity was
10 poured onto each plate. The assay was performed as described previously (Kopp et al., 1993) with the following modifications: To prepare 100 ml of final volume of the overlay assay mix, 3.4 g of trehalose was dissolved in 80 ml of 200 mM citric acid, pH 4.5, 5 mM EDTA; 1g of agarose was added, and the mixture was melted in a microwave oven and then cooled to 50°C. Immediately before pouring
15 the mixture onto the replica-plated colonies, 2 ml of *N*-ethylmaleimide (2.5 mg/ml), 985 units of horseradish peroxidase (EC 1.11.1.7), 800 units of glucose oxidase (EC 1.1.3.4), and 4.8 ml of *o*-dianisidine (10 mg/ml) were added. The overlay-assay mixture was incubated on the plates for 15 minutes at 24°C. Colonies with secreted acid trehalase activity developed a dark green color whereas
20 the other transformants remained white.

Assays: liquid trehalase assays were performed as described previously (Kopp et al., 1993). Proteinase A was assayed according to Wiemken et al. (Wiemken et al., 1979), using denatured hemoglobin.

Amplification of Plasmid Library: a YEp24 genomic plasmid library
25 (Carlson and Botstein, 1982) was kindly provided by D. Botstein (Stanford University). Amplification of the library was performed as described previously (Kopp et al., 1993).

Cloning, Sequencing and DNA Analysis of *ATH1*: genomic and plasmid DNA from *S. cerevisiae* and plasmid DNA from *E. coli* were prepared as
30 described previously (Birnboim and Doly, 1979; Sherman et al., 1979). Standard procedures were followed for subcloning DNA fragments and for identifying

recombinant clones (Maniatis et al., 1982). After screening for secreted acid trehalase activity, plasmid DNA from the positive-reacting yeast transformant colonies was isolated (Sherman et al., 1979); plasmids containing 8.5 and 10-kbp inserts were recovered. To determine the nucleotide sequence, *SaII* and *EcoRI* restriction fragments from the 8.5-kbp insert were subcloned into the vectors pTZ18R and pTZ19R (Pharmacia, Freiburg, Germany) to construct plasmids pMATZ1 to pMATZ4. The nucleotide sequence was determined by the dideoxy chain termination method (Sanger et al., 1977). The sequence containing the *ATH1* gene was determined on both the coding and noncoding strands.

10 The 0.3 kb *EcoRI/SaII* fragment from pDAT1.9 was used as a probe in Southern (Southern, 1975) and Northern blot analyses. Radiolabeled DNA hybridization probes were prepared by the random priming method (Feinberg and Vogelstein, 1983). For Southern blot analysis, the genomic DNA was digested with *EcoRI*, separated on a 0.8% agarose gel, incubated in 0.25 M HCl, and blotted onto a Hybond N⁺ membrane in 0.4 M NaOH. For Northern blot analysis, the RNA was prepared by the method of Chirgwin (Chirgwin et al., 1979). Following electrophoresis, the RNA was transferred to a Hybond N⁺ membrane in 0.04 M NaOH.

20 Deletion of *ATH1*: Plasmid pMATZ1 contains two *EcoRV* sites, one in the 5' non-coding region and the other within the open reading frame (Figure 4). This plasmid was restricted with *EcoRV* to remove a 2.4-kbp fragment encoding most of the open reading frame of *ATH1*. The 1.1-kbp *HindIII* fragment containing the *URA3* gene was isolated from plasmid YEp24, and the overhanging 5'-ends were filled in by treatment with the Klenow fragment of DNA polymerase I. The blunt-ended *URA3* fragment was cloned into pMATZ1 which had been digested with *EcoRV* to generate plasmid pMATZ1.1. The *EcoRI* fragment from plasmid pMATZ1.1 was isolated and used to transform yeast strain SEY6210 with approximately 10 μ g of DNA. Ura⁺ colonies were isolated and examined by Southern blotting to confirm the site of integration. Yeast strain MDY3 contained the *URA3* gene integrated at the chromosomal *ATH1* locus.

25

30

Identification of two ATH-secreting
gene encoding the vacuolar acid trehalase,
approaches. First, peptide sequences were obtained from a
protein fraction with high acid trehalase activity (Mittenbühler and Holzer, 1988).
5 Degenerate oligonucleotides were synthesized based on the peptide sequences and
were used in a polymerase chain reaction with genomic DNA from the yeast strain
(Destruelle et al., 1994). Cloning of the corresponding gene led to the isolation
of a novel yeast gene, *YGPI* (Destruelle et al., 1994). The *YGPI* gene codes for
a highly glycosylated, secreted protein with an unknown function. The *YGPI*
10 gene, however, is not the structural gene for acid trehalase. It appears that
additional proteins may have been retained during the initial purification
(Mittenbühler and Holzer, 1988) of acid trehalase.

In the second approach, we relied on the observation that overproduction
of vacuolar proteins can lead to their expression at the cell surface (Rothman et al.,
15 1986; Stevens et al., 1986). Secreted proteins can then be identified by
immunoblotting with a specific antibody or by their enzymatic activity. For the
cloning of acid trehalase, we developed a specific enzymatic overlay assay that
allowed the convenient screening of many transformants (see Materials and
Methods). The activity of neutral trehalase was inhibited by the addition of 5 mM
20 EDTA and the acidic pH of 4.5, where the enzyme shows very little activity (App
and Holzer, 1989). To identify putative ATH-encoding clones, yeast strain
SEY6210 was transformed with plasmid DNA from a YEp24-based genomic
library (Carlson and Botstein, 1982). Between 200 and 500 transformant colonies
per plate were replica-plated on YNB-plates containing fructose as a carbon source
25 and assayed for secreted acid trehalase activity. In a screen of approximately
10,000 Ura⁺ transformants, nine were positive for secreted ATH activity.
Reintroducing the purified plasmids (pDAT1.1-pDAT1.9) into yeast resulted in the
secretion of acid trehalase activity. Thus, pDAT1.1-pDAT1.9 carry DNA
sequences that cause yeast cells to secrete a catalytically active portion of their acid
30 trehalase.

of the DNA fragment that confers an ATH secretion phenotype: mapping of the plasmid DNA isolated from the yeast cells revealed two plasmids with overlapping genomic inserts of 10 and 8.5-kbp. The plasmids were named pDAT1.8 and pDAT1.9, respectively. To define the limits of the DNA segment leading to acid trehalase secretion, various subclones of pDAT1.9 were constructed in plasmids YEp24 and pSEY8 (Emr et al., 1986), and yeast transformants carrying the subcloned plasmids were examined for secretion of ATH. None of the subclones showed secreted acid trehalase activity indicating that the functional gene sequence was larger than that contained on any of the subcloned fragments. Therefore, the entire nucleotide sequence of the 8.5-kbp insert was determined by sequence analysis. For that purpose, the *EcoRI* and *SaII*-fragments from the genomic insert of pDAT1.9 were subcloned into the sequencing vectors pTZ18R and pTZ19R. Analysis of the sequence revealed two open reading frames from which one has been described previously as part of the *YMNI* gene (Thorsness et al., 1993). The second open reading frame (SEQUENCE ID NO:1), which showed no homology to sequences in the EMBL and GenBank nucleotide libraries, was 3,126 bp and is contained in the *EcoRI* fragment from pDAT1.9.7. The 5' noncoding region contains two possible TATA boxes. The open reading frame encodes a 1,041 amino acid protein (SEQUENCE ID NO:2) with a predicted molecular mass of approximately 117,400 Da. The coding region contains 25 potential N-glycosylation sites. The gene was named *ATH1* (Accession Number: X84156 *S. cerevisiae* *ATH1* gene), for yeast acid trehalase.

Characterization of *ATH1*: The amino acid sequence deduced from the *ATH1* gene was compared with those of proteins in the SWISS-Prot and PIR protein databases by use of the FASTA algorithm and the Wordsearch program of the University of Wisconsin Genetics Computer Group package (Devereux et al., 1984). This analysis did not reveal any homology to the five cloned trehalases from different organisms (Gutierrez et al., 1989; Kopp et al., 1993; Ruf et al., 1990; Su et al., 1993; Takiguchi et al., 1992) nor to any other protein in the libraries. *Ath1p* lacks a characteristic signal sequence at the amino terminus as expected for a soluble secretory pathway protein. In addition, there are no hydrophobic

- domains that are likely to serve as internal signal sequences. The deduced amino acid sequence also does not reveal a consensus signal sequence cleavage site based on the rules of von Heijne (von Heijne, 1986); there are no positive S values indicating likely cleavage sites within the first N-terminal 300 amino acids.
- 5 However, localization of Ath1p to the vacuole could occur by a mechanism independent of the secretory pathway (Klionsky et al., 1992). ATH has been characterized as a glycosylated protein that transits to the vacuole in a *sec*-dependent manner, however, suggesting movement through the secretory pathway (Londesborough and Varimo, 1984; Harris and Cotter, 1988; Mittenbühler and
- 10 Holzer, 1988).

To confirm the requirement of the *ATH1* gene for acid trehalase activity, we carried out a one step gene transplacement (Rothstein, 1983). The *ATH1* gene was disrupted at the chromosomal locus to generate the mutant yeast strain MDY3. The mutant strain has no detectable acid trehalase activity as determined using the

15 overlay assay or liquid assays with crude cell extracts (Table 1).

Table 1. Enzymatic activities of vacuolar and cytosolic proteins in wild type strains and a strain overexpressing the *ATH1* gene.

5	Strain	ATH activity (mU/mg)	NTH activity (mU/mg)	Alkaline phosphatase activity (mU/mg)	Glucose-6-phosphate Proteinase	
					A dehydrogenase activity (mU/mg)	
	SEY6210	8.5	19.5	100	8.6160	
	SEY6210/YEp24	6.5	20.1	60	7.5175	
	SEY6210/pDAT1.9	67.0	26.1	70	7.6200	14

These results indicate that the *ATH1* gene product is required for ATH activity but do not demonstrate whether *ATH1* is the structural gene for acid trehalase or encodes a regulatory protein.

Northern blot analysis revealed that *ATH1* is expressed in stationary phase
5 cells while no expression could be detected in logarithmically growing yeast cells. The expression pattern corresponds with the activity profile of acid trehalase. To further investigate if *ATH1* is the structural gene for acid trehalase or a putative regulator, activities of different vacuolar and cytosolic enzymes were measured in the acid trehalase-overproducing strain, the mutant strain MDY3 and in a wild type
10 strain. As shown in Table 1, cells containing pDAT1.9 exhibit about an 8-10 fold higher level of acid trehalase activity than the same strain carrying the parent 2 μ plasmid YEp24. Of the enzyme activities examined, only acid trehalase is dramatically increased in cells containing pDAT1.9; the activities of other vacuolar proteins (alkaline phosphatase and proteinase A) and a cytosolic protein (glucose-6-
15 phosphate dehydrogenase) are not elevated. The activity of neutral trehalase is slightly increased upon overproduction of *ATH1*. In the $\Delta ath1$ strain, however, ATH activity is completely eliminated while there is no effect on NTH activity. The enzymatic activities of acid and neutral trehalases in the $\Delta ath1$ strain are at the same levels as are seen in an acid trehalase mutant generated by random
20 mutagenesis with ethyl methane sulfonate (Destruelle, 1993); the mutation completely eliminates ATH activity while having no effect on the activity of NTH. The plasmid pDAT1.9 complements the mutagen-induced defect but is not able to complement a mutant lacking neutral trehalase activity (Destruelle, 1993).

25 References

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30 All publications and patent applications cited in this specification are herein incorporated by reference as if each individual publication or patent application

were specifically and individually indicated to be incorporated by reference. Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be readily apparent to those of ordinary skill in the art in light of the teachings of this
5 invention that certain changes and modifications may be made thereto without departing from the spirit or scope of the appended claims.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: The Regents of the University
of California
- (ii) TITLE OF INVENTION: STRESS TOLERANT YEAST MUTANTS
- (iii) NUMBER OF SEQUENCES: 2
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: ROBBINS, BERLINER & CARSON
 - (B) STREET: 201 N. Figueroa Street, 5th Floor
 - (C) CITY: Los Angeles
 - (D) STATE: California
 - (E) COUNTRY: USA
 - (F) ZIP: 90012-2628
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: US
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
- (vii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Berliner, Robert
 - (B) REGISTRATION NUMBER: 20,121
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 - (B) TELEFAX: (213) 977-1003
 - (C) TELEX:

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 3876 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 579..3701

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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TCCTTTATGA GGCCTGCTAG TATCACCTAA TATTGCATCT GTTTTACCAG AAATTCCTC	180
ATTCCAATAA TGAAAAGAAT AAGATCGCTT TGGTTCAATG CGGAGGCTTC TTATTCAAAC	240
CTTAACAATT CTCCTAGTTT GAGGAACAAA AATAGTACCG GTAATAACTC TCGCTCTAAA	300
AATTATCGTT CTTTCTCAAG GTTTGACTTG ATCAACTCTA TACTTTTACT GATGATGCTA	360
TTTTTATTAG CTATCTTCGT CACTGCATTA TATTTAACAA AGCGTTCCAG GCTTACATAC	420

TCACATGCCT CGAGGGCTGC CCTATTTAAC CTCTGGGTGT GATATCGCCG TCATTGGGAA	480
ATCATACGTT GAACTACGAT CCAGAAGCAA GGGAACTCTT TAAAAAACTT TATGAACTCC	540
TTTCTGATTT CAACACGGCA TATTATGATG ATGAGAAC ATG ATT TTG GGA AGT	593
Met Ile Leu Gly Ser	
1 5	
AAC TTG TTC TCA AAG AAT ACA TAC TCG AGA CAA CCA TAT GTT GCT AAC	641
Asn Leu Phe Ser Lys Asn Thr Tyr Ser Arg Gln Pro Tyr Val Ala Asn	
10 15 20	
GGT TAT ATA GGT AGT CGT ATT CCC AAT ATT GGG TTC GGC TAT GCC TTA	689
Gly Tyr Ile Gly Ser Arg Ile Pro Asn Ile Gly Phe Gly Tyr Ala Leu	
25 30 35	
GAC ACC CTG AAT TTT TAC ACA GAC GCA CCA GGC GCT TTG AAT AAC GGT	737
Asp Thr Leu Asn Phe Tyr Thr Asp Ala Pro Gly Ala Leu Asn Asn Gly	
40 45 50	
TGG CCC TTA AGA AAT CAT AGA TTT GCC GGT GCG TTT GTA TCG GAC TTT	785
Trp Pro Leu Arg Asn His Arg Phe Ala Gly Ala Phe Val Ser Asp Phe	
55 60 65	
TAT TGT CTA CAA CCA AAA CTA AAT TCA ACA AAC TTC CCA GAA TTG GAT	833
Tyr Cys Leu Gln Pro Lys Leu Asn Ser Thr Asn Phe Pro Glu Leu Asp	
70 75 80 85	
GAT GTA GGA TAT TCC ACT GTC ATT TCA TCT ATT CCA CAA TGG ACC AAT	881
Asp Val Gly Tyr Ser Thr Val Ile Ser Ser Ile Pro Gln Trp Thr Asn	
90 95 100	
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Leu Gln Phe Ser Leu Val Asn Asp Ser Lys Trp Phe Asn Pro Gln Asn	
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GTT ACG TTG GAT GAC GTA ACT AAT TAT AGC CAA AAC TTA TCA ATG AAG	977
Val Thr Leu Asp Asp Val Thr Asn Tyr Ser Gln Asn Leu Ser Met Lys	
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Asp Gly Ile Val Thr Thr Glu Leu Asp Trp Leu Asn Ser Gln Ile His	
135 140 145	
GTT AAA AGT GAA ATC TGG GCA CAT CGG CAC ATT CAT CCA CTG GGA GTG	1073
Val Lys Ser Glu Ile Trp Ala His Arg His Ile His Pro Leu Gly Val	
150 155 160 165	
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Val Ser Leu Glu Ile Ser Leu Asn Thr Asp His Leu Pro Ser Asp Phe	
170 175 180	
GAT TCA TTA GAT GTT AAT ATA TGG GAT ATA CTT GAT TTC AAC ACA TCA	1169
Asp Ser Leu Asp Val Asn Ile Trp Asp Ile Leu Asp Phe Asn Thr Ser	
185 190 195	
CAT AGG ACT GTT CTA CAT AGC ACG GGA ACA GAC GAA AAA AAT AAT GCG	1217
His Arg Thr Val Leu His Ser Thr Gly Thr Asp Glu Lys Asn Asn Ala	
200 205 210	
GTT TTC ATG ATT GTT CAG CCA GAT AAC GTT CCA TCT TCT AAT TGC GCT	1265
Val Phe Met Ile Val Gln Pro Asp Asn Val Pro Ser Ser Asn Cys Ala	
215 220 225	
ATT TAC TCA ACG TGT ACT GTA AAG TAT GAA AAT TCC ACC AAT CCA ATA	1313
Ile Tyr Ser Thr Cys Thr Val Lys Tyr Glu Asn Ser Thr Asn Pro Ile	
230 235 240 245	
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Asn Ser Ser Glu Ser Phe Glu Glu Lys Asp Val Ser Ser Asn Ile Tyr	
250 255 260	

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GGG ATT GAT GAC GAG TAT CTG AGA TAT ACT ACA TGG CCA ATT ATC AAA Gly Ile Asp Asp Glu Tyr Leu Arg Tyr Thr Trp Pro Ile Ile Lys 470 475 480 485	2033
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CAC ATC AAT AAC GGG GCT TTC ACG AAT GCT GGC ATT AAA ACA CTT CTA His Ile Asn Asn Gly Ala Phe Thr Asn Ala Gly Ile Lys Thr Leu Leu 520 525 530	2177
AAG TGG GCA ACA GAC ATT GGC AAT CAT CTC GGC GAG GTC GTT GAC CCC Lys Trp Ala Thr Asp Ile Gly Asn His Leu Gly Glu Val Val Asp Pro 535 540 545	2225

AAA TGG AGT GAA ATT TCC AAA GAT ATT TAT ATC CCT AGA TCC TCA TCT Lys Trp Ser Glu Ile Ser Lys Asp Ile Tyr Ile Pro Arg Ser Ser Ser 550 555 560 565	2273
AAC ATC ACT TTG GAA TAT TCT GGT ATG AAT AGC TCA GTG GAG ATT AAA Asn Ile Thr Leu Glu Tyr Ser Gly Met Asn Ser Ser Val Glu Ile Lys 570 575 580	2321
CAG GCG GAT GTG ACT TTA ATG GTT TAC CCA CTT GGA TAT ATC AAT GAT Gln Ala Asp Val Thr Leu Met Val Tyr Pro Leu Gly Tyr Ile Asn Asp 585 590 595	2369
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22

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(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1041 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

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1				5					10					15	

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 Phe Gly Tyr Ala Leu Asp Thr Leu Asn Phe Tyr Thr Asp Ala Pro Gly
 35 40 45
 Ala Leu Asn Asn Gly Trp Pro Leu Arg Asn His Arg Phe Ala Gly Ala
 50 55 60
 Phe Val Ser Asp Phe Tyr Cys Leu Gln Pro Lys Leu Asn Ser Thr Asn
 65 70 75 80
 Phe Pro Glu Leu Asp Asp Val Gly Tyr Ser Thr Val Ile Ser Ser Ile
 85 90 95
 Pro Gln Trp Thr Asn Leu Gln Phe Ser Leu Val Asn Asp Ser Lys Trp
 100 105 110
 Phe Asn Pro Gln Asn Val Thr Leu Asp Asp Val Thr Asn Tyr Ser Gln
 115 120 125
 Asn Leu Ser Met Lys Asp Gly Ile Val Thr Thr Glu Leu Asp Trp Leu
 130 135 140
 Asn Ser Gln Ile His Val Lys Ser Glu Ile Trp Ala His Arg His Ile
 145 150 155 160
 His Pro Leu Gly Val Val Ser Leu Glu Ile Ser Leu Asn Thr Asp His
 165 170 175
 Leu Pro Ser Asp Phe Asp Ser Leu Asp Val Asn Ile Trp Asp Ile Leu
 180 185 190
 Asp Phe Asn Thr Ser His Arg Thr Val Leu His Ser Thr Gly Thr Asp
 195 200 205
 Glu Lys Asn Asn Ala Val Phe Met Ile Val Gln Pro Asp Asn Val Pro
 210 215 220
 Ser Ser Asn Cys Ala Ile Tyr Ser Thr Cys Thr Val Lys Tyr Glu Asn
 225 230 235 240
 Ser Thr Asn Pro Ile Asn Ser Ser Glu Ser Phe Glu Glu Lys Asp Val
 245 250 255
 Ser Ser Asn Ile Tyr Asn Val Ile Leu Arg Glu Asp Gln Pro Lys Ile
 260 265 270
 Ile Val His Lys Tyr Val Gly Ile Met Ser Thr Glu Phe Asn Lys Asn
 275 280 285
 Lys Glu Gln Gln Asp Asn Thr Asn Ile Gly Leu Ala Lys Met Ile Ala
 290 295 300
 Leu Asn Ser Lys Gly Asn Tyr Glu Lys Leu Leu Ser Ser His Lys Arg
 305 310 315 320
 Ala Trp Tyr Asp Leu Tyr Asn Asp Ala Phe Ile Glu Ile Pro Ser Asp
 325 330 335
 Ser Leu Leu Glu Met Thr Ala Arg Ser Ser Leu Phe His Leu Leu Ala
 340 345 350
 Asn Thr Arg Asp Tyr Asn Val Ser Ser Asp Arg Gly Leu Pro Val Gly
 355 360 365
 Val Ser Gly Leu Ser Ser Asp Ser Tyr Gly Gly Met Val Phe Trp Asp
 370 375 380
 Ala Asp Ile Trp Met Glu Pro Ala Leu Leu Pro Phe Phe Pro Asn Val
 385 390 395 400

24

Ala Gln Asn Met Asn Asn Tyr Arg Asn Ala Thr His Ser Gln Ala Lys
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 Leu Asn Ala Glu Lys Tyr Gly Tyr Pro Gly Ala Ile Tyr Pro Trp Thr
 420 425 430
 Ser Gly Lys Tyr Ala Asn Cys Thr Ser Thr Gly Pro Cys Val Asp Tyr
 435 440 445
 Glu Tyr His Ile Asn Val Asp Val Ala Met Ala Ser Phe Ser Ile Tyr
 450 455 460
 Leu Asn Gly His Glu Gly Ile Asp Asp Glu Tyr Leu Arg Tyr Thr Thr
 465 470 475 480
 Trp Pro Ile Ile Lys Asn Ala Ala Gln Phe Phe Thr Ala Tyr Val Lys
 485 490 495
 Tyr Asn Ser Ser Leu Gly Leu Tyr Glu Thr Tyr Asn Leu Thr Asp Pro
 500 505 510
 Asp Glu Phe Ala Asn His Ile Asn Asn Gly Ala Phe Thr Asn Ala Gly
 515 520 525
 Ile Lys Thr Leu Leu Lys Trp Ala Thr Asp Ile Gly Asn His Leu Gly
 530 535 540
 Glu Val Val Asp Pro Lys Trp Ser Glu Ile Ser Lys Asp Ile Tyr Ile
 545 550 555 560
 Pro Arg Ser Ser Ser Asn Ile Thr Leu Glu Tyr Ser Gly Met Asn Ser
 565 570 575
 Ser Val Glu Ile Lys Gln Ala Asp Val Thr Leu Met Val Tyr Pro Leu
 580 585 590
 Gly Tyr Ile Asn Asp Glu Ser Ile Leu Asn Asn Ala Ile Lys Asp Leu
 595 600 605
 Tyr Tyr Tyr Ser Glu Arg Gln Ser Ala Ser Gly Pro Ala Met Thr Tyr
 610 615 620
 Pro Val Phe Val Ala Ala Ala Ala Gly Leu Leu Asn His Gly Ser Ser
 625 630 635 640
 Ser Gln Ser Tyr Leu Tyr Lys Ser Val Leu Pro Tyr Leu Arg Ala Pro
 645 650 655
 Phe Ala Gln Phe Ser Glu Gln Ser Asp Asp Asn Phe Leu Thr Asn Gly
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 Leu Thr Gln Pro Ala Phe Pro Phe Leu Thr Ala Asn Gly Gly Phe Leu
 675 680 685
 Gln Ser Ile Leu Phe Gly Leu Thr Gly Ile Arg Tyr Ser Tyr Glu Val
 690 695 700
 Asp Pro Asp Thr Lys Lys Ile Asn Arg Leu Leu Arg Phe Asn Pro Ile
 705 710 715 720
 Glu Leu Pro Leu Leu Pro Gly Gly Ile Ala Ile Arg Asn Phe Lys Tyr
 725 730 735
 Met Asn Pro Val Leu Asp Ile Ile Ile Asp Asp His Asn Gly Thr Ile
 740 745 750
 Val His Lys Ser Gly Asp Val Pro Ile His Ile Lys Ile Pro Asn Arg
 755 760 765
 Ser Leu Ile His Asp Gln Asp Ile Asn Phe Tyr Asn Gly Ser Glu Asn
 770 775 780

25

Glu Arg Lys Pro Asn Leu Glu Arg Arg Asp Val Asp Arg Val Gly Asp
 785 790 795 800
 Pro Met Arg Met Asp Arg Tyr Gly Thr Tyr Tyr Leu Leu Lys Pro Lys
 805 810 815
 Gln Glu Leu Thr Val Gln Leu Phe Lys Pro Gly Leu Asn Ala Arg Asn
 820 825 830
 Asn Ile Ala Glu Asn Lys Gln Ile Thr Asn Leu Thr Ala Gly Val Pro
 835 840 845
 Gly Asp Val Ala Phe Ser Ala Leu Asp Gly Asn Asn Tyr Thr His Trp
 850 855 860
 Gln Pro Leu Asp Lys Ile His Arg Ala Lys Leu Leu Ile Asp Leu Gly
 865 870 875 880
 Glu Tyr Asn Glu Lys Glu Ile Thr Lys Gly Met Ile Leu Trp Gly Gln
 885 890 895
 Arg Pro Ala Lys Asn Ile Ser Ile Ser Ile Leu Pro His Ser Glu Lys
 900 905 910
 Val Glu Asn Leu Phe Ala Asn Val Thr Glu Ile Met Gln Asn Ser Gly
 915 920 925
 Asn Asp Gln Leu Leu Asn Glu Thr Ile Gly Gln Leu Leu Asp Asn Ala
 930 935 940
 Gly Ile Pro Val Glu Asn Val Ile Asp Phe Asp Gly Ile Glu Gln Glu
 945 950 955 960
 Asp Asp Glu Ser Leu Asp Asp Val Gln Ala Leu Leu His Trp Lys Lys
 965 970 975
 Glu Asp Leu Ala Lys Leu Ile Asp Gln Ile Pro Arg Leu Asn Phe Leu
 980 985 990
 Lys Arg Lys Phe Val Lys Ile Leu Asp Asn Val Pro Val Ser Pro Ser
 995 1000 1005
 Glu Pro Tyr Tyr Glu Ala Ser Arg Asn Gln Ser Leu Ile Glu Ile Leu
 1010 1015 1020
 Pro Ser Asn Arg Thr Thr Phe Thr Ile Asp Tyr Asp Lys Phe Ala Gly
 1025 1030 1035 1040
 Gly

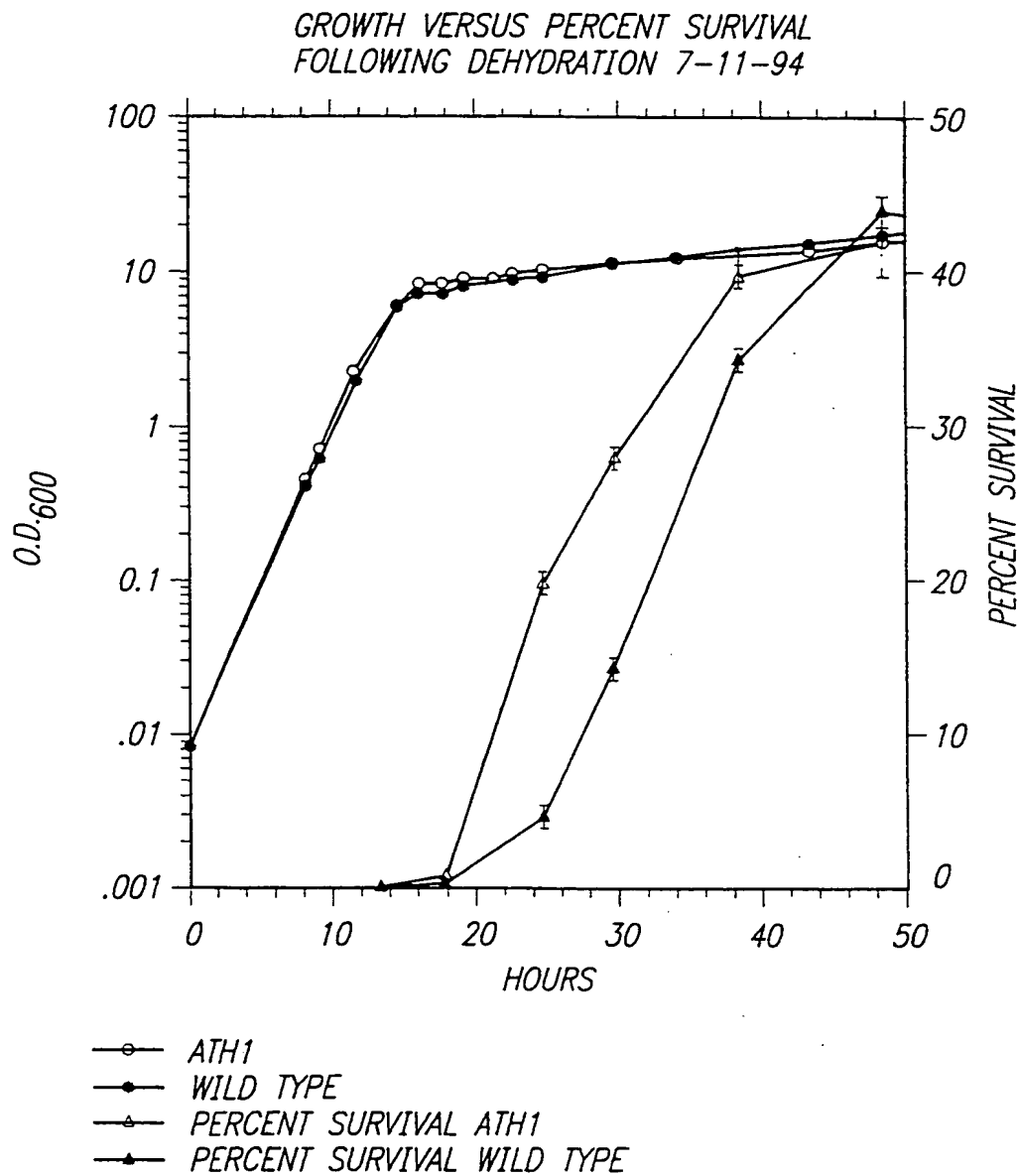
WHAT IS CLAIMED IS:

1. An isolated yeast mutant deficient in the expression of functional Ath1p gene product.
2. An isolated yeast according to claim 1, wherein said mutant or an ancestor of said mutant was generated by genetically engineering a yeast cell to create a mutation in an Ath1p allele of said yeast cell.
3. An isolated yeast according to claim 1, wherein said mutant expresses less than 10% of that expressed by the corresponding wild-type yeast.
4. An isolated yeast mutant according to claim 1, as deposited in the ATCC deposit # _____.
5. An isolated nucleic acid comprising ATH1 (SEQUENCE ID NO:1) or fragment thereof capable of hybridizing under stringent conditions with ATH1.
6. An isolated nucleic acid according to claim 5 comprising in 5' - 3' orientation, a first ATH1 fragment thereof capable of hybridizing under stringent conditions with ATH1, an intervening sequence, and a second different ATH1 fragment thereof capable of hybridizing under stringent conditions with ATH1.
7. A method for producing a yeast mutant with improved survival ability under stress conditions, said method comprising steps:
 - subjecting a population of yeast to stress conditions;
 - detecting in said population a yeast mutant deficient in the expression of functional Ath1p gene product;
 - isolating said yeast mutant; and,
 - growing said yeast mutant to obtain yeast with improved survival ability under stress conditions.

8. A method according to claim 7, said detecting step comprises contacting said population with an Ath1p gene product-specific reagent.
9. A method according to claim 7, said detecting step comprising contacting said population with an isolated nucleic acid comprising ATH1 (SEQUENCE ID NO:1) or fragment thereof capable of hybridizing under stringent conditions with ATH1.
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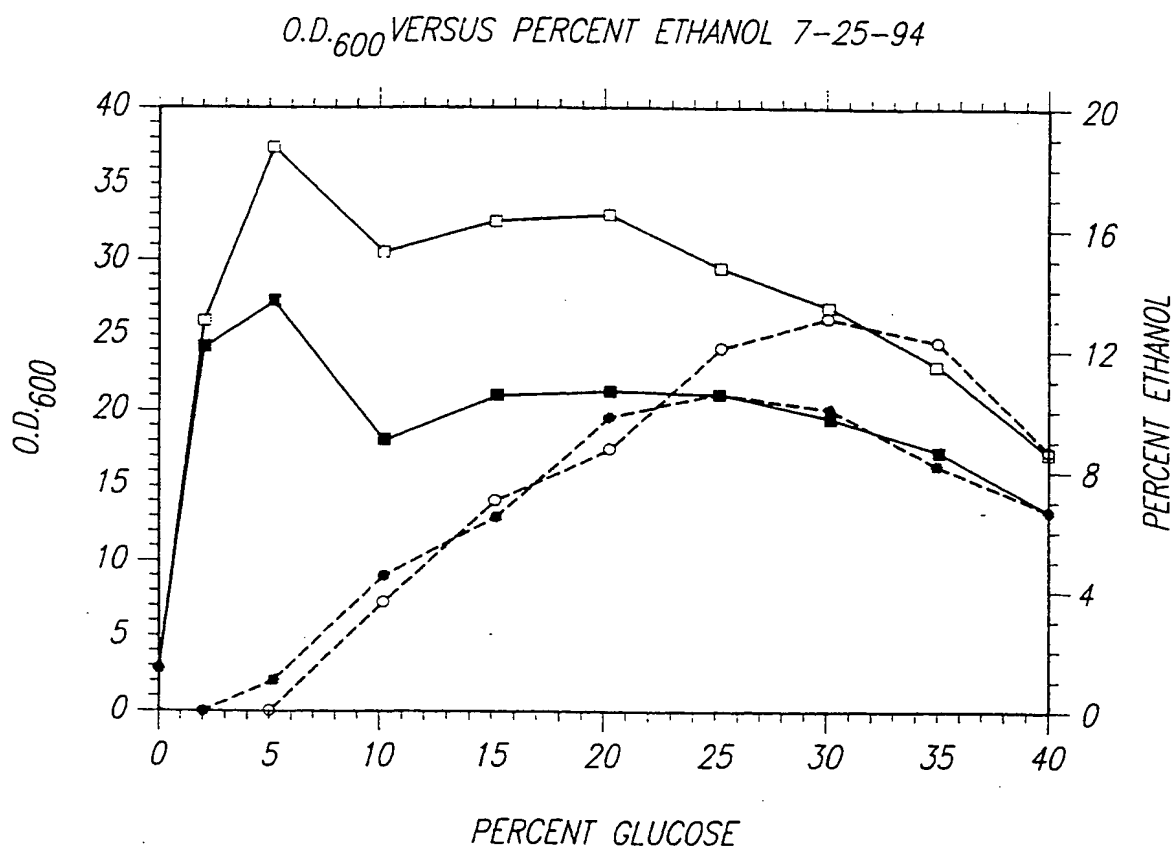
FIG. 1



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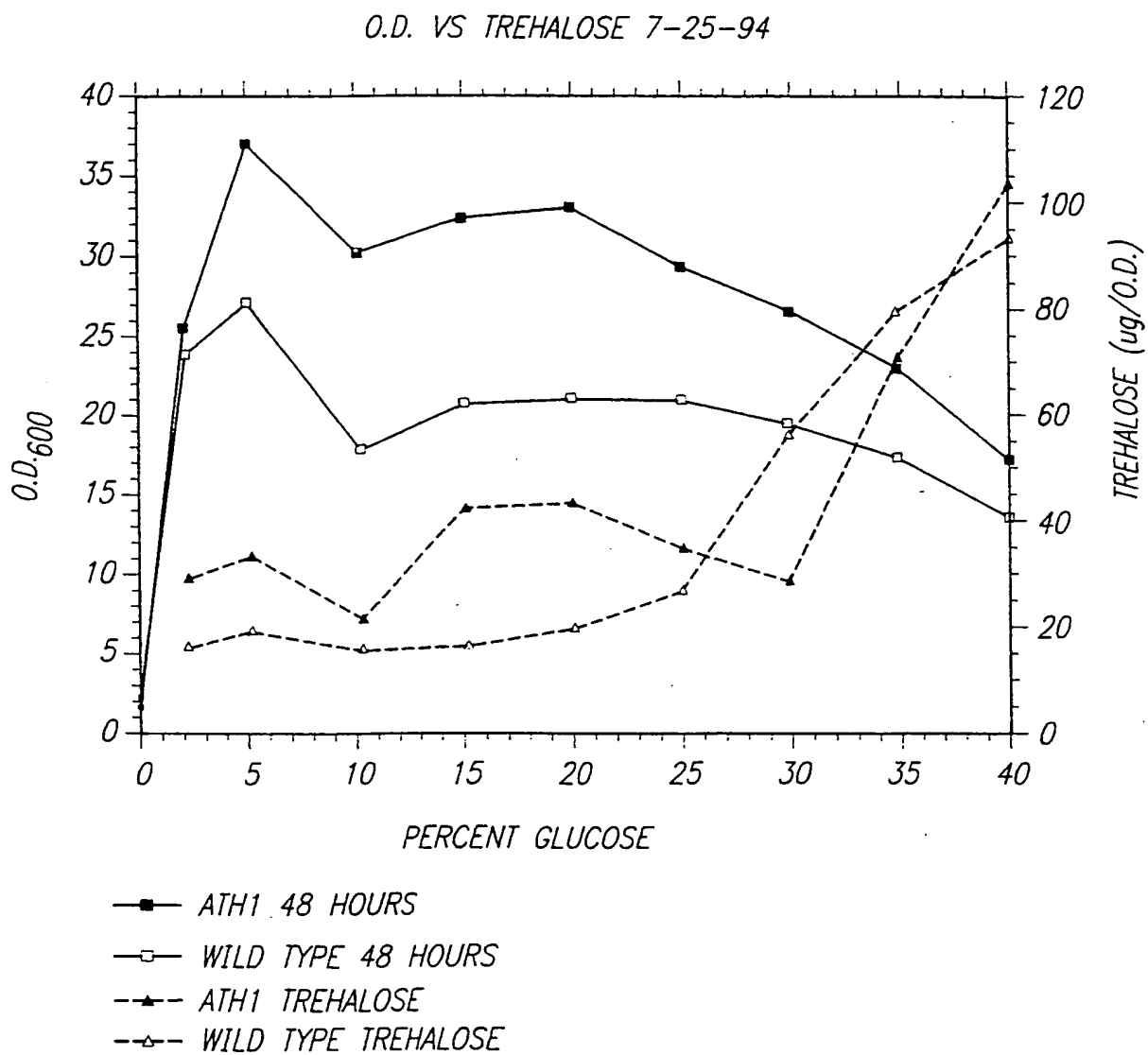
FIG. 2



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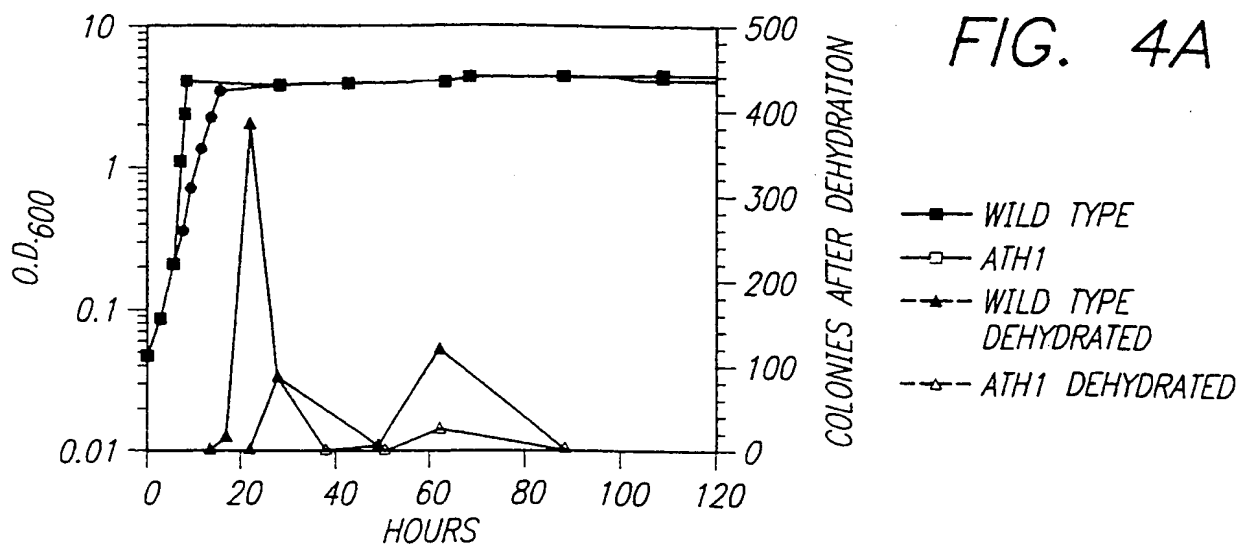
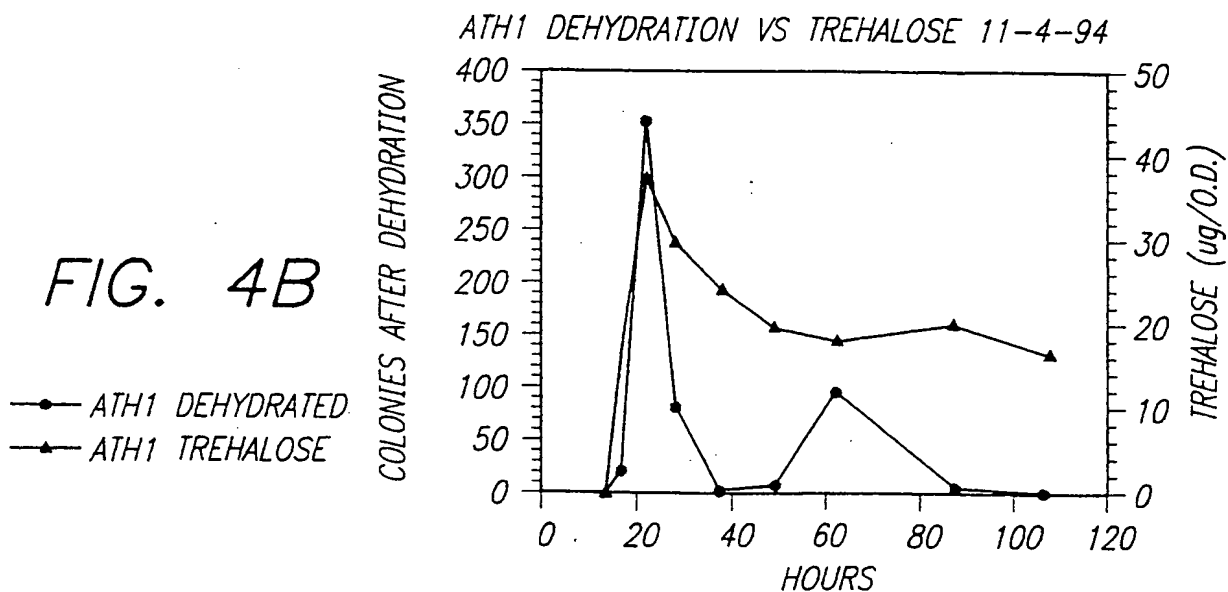
FIG. 3



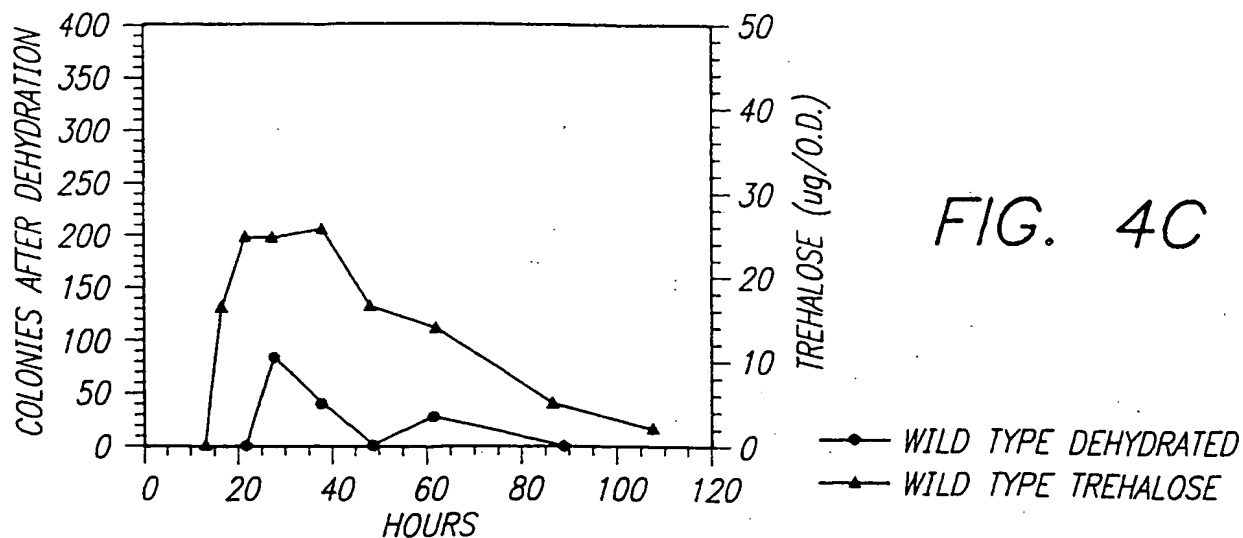
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O.D. VS DEHYDRATION WITH STANDARD DEVIATION 11-4-94

**FIG. 4B**

WILD TYPE DEHYDRATION vs TREHALOSE 11-4-94



INTERNATIONAL SEARCH REPORT

International application No.
PCT/US96/10782

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : Please See Extra Sheet.

US CL : Please See Extra Sheet.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : Please See Extra Sheet.

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched
Please See Extra Sheet.Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
Please See Extra Sheet.

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	FEBS Letters, Volume 360, issued 1995, Nwaka et al., "Phenotypic features of trehalase mutants in <i>Saccharomyces cerevisiae</i> ", pages 286-290, see entire document.	1-4



Further documents are listed in the continuation of Box C.



See patent family annex.

* Special categories of cited documents:	*T	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
A document defining the general state of the art which is not considered to be of particular relevance	*X*	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
E earlier document published on or after the international filing date	*Y*	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
L document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*Z*	document member of the same patent family
O document referring to an oral disclosure, use, exhibition or other means		
P document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search

30 JULY 1996

Date of mailing of the international search report

17 SEP 1996

Name and mailing address of the ISA/US
Commissioner of Patents and Trademarks
Box PCT
Washington, D.C. 20231

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INTERNATIONAL SEARCH REPORT

International application No.
PCT/US96/10782

A. CLASSIFICATION OF SUBJECT MATTER:
IPC (6):

C12N 1/15, 15/01, 15/31

A. CLASSIFICATION OF SUBJECT MATTER:
US CL :

435/6, 254.2, 254.21; 536/23.1

B. FIELDS SEARCHED

Minimum documentation searched

Classification System: U.S.

435/6, 254.2, 254.21; 536/23.1

B. FIELDS SEARCHED

Documentation other than minimum documentation that are included in the fields searched:

NONE

B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

MEDLINE, APS

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